Histopathological Change of Oral Malignant Tumour and Epithelial Dysplasia Subjected to Photodynamic Therapy

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ABSTRACT

Objectives: The purpose of this study is to analyze the morphological change of cell nuclei and the change of proliferating activity of oral malignancy and epithelial dysplasia between before and after photodynamic therapy in order to predict recurrence.

Material and Methods: We experienced 14 cases of oral squamous cell carcinoma, one case of verrucous carcinoma and seven cases of epithelial dysplasia treated by photodynamic therapy (PDT). The mean nuclear area (NA) and coefficient of variation of the nuclear area (NACV) of 100 nuclei per slide were calculated using computer-assisted image analysis in hematoxylin and eosin stained biopsy specimens before and after PDT. Additionally, proliferating cell nuclear antigen (PCNA) immunohistochemistry was carried out in each specimen.

Results: The mean NA after PDT was significantly lower than that before PDT in the nonrecurrent group. However, there was no significant difference in mean NA before and after PDT in the recurrent group. There were no significance differences in NACV before and after PDT in either the nonrecurrent or recurrent group. Furthermore, the PCNA labelling indices of the specimens after PDT was significantly lower than that before PDT in both the nonrecurrent and the recurrent group.

Conclusions: Mean nuclear area in the biopsy specimen after photodynamic therapy is likely to be a predictive marker for the recurrence of oral squamous cell carcinoma or epithelial dysplasia subjected to photodynamic therapy, while coefficient of variation of the nuclear area and proliferating cell nuclear antigen labelling indices are less helpful in predicting the recurrence of such lesions.

Keywords: photodynamic therapy; squamous cell carcinoma; epithelial cells; carcinoma, verrucous; cell nucleus; proliferating cell nuclear antigen.
INTRODUCTION

Photodynamic therapy (PDT) consists of the administration of a photosensitizer that localizes in malignant or premalignant lesions because of its relative selectivity, followed by light irradiation. PDT uses the localized delivery of light to activate the photosensitizer \[1,2\]. As PDT is a useful therapy for oral lesions because of the accessibility of light to the lesions, it has been applied to oral malignant and premalignant lesions \[3-6\]. However, the recurrence of tumours after PDT despite a good initial response remains problematic \[7-9\]. In a previous study, we pointed out the heterogeneous susceptibility of mouse tumours to PDT anti-tumour effects \[10\]. Furthermore, we also found that the nuclear area (NA) and coefficient of variation of the nuclear area (NACV) as well as proliferating cell nuclear antigen (PCNA) of tumour cells are useful indicators in assessing the effect on tumour of PDT \[11,12\].

We present here 14 cases of oral squamous cell carcinoma (SCC), one case of verrucous carcinoma and 7 cases of oral epithelial dysplasia (ED), which were clinically diagnosed as leukoplakia, treated by PDT. No biopsy, taken from 4 to 9 weeks after PDT, produced evidence of residual tumour. However, 5 cases of SCC and one case of ED recurred 4 to 30 months after PDT. The biopsy specimens before and after PDT were investigated using morphological analyses of the tumour and ED cell nuclei as well as PCNA immunohistochemistry in order to clarify the correlation between the histopathological changes of lesions after PDT and during recurrence.

MATERIAL AND METHODS

Patients and PDT

Twenty patients were diagnosed with 22 lesions having one or more histologically proven SCC, verrucous carcinoma or ED of the oral cavity (Table 1). All lesions were diagnosed as stage I or II (T1 or T2 N0M0) SCC, verrucous carcinoma or ED, and no deep extension to underlying muscle or bone was identified by the clinical, histological or radiological findings. None of the patients had received prior treatment. Informed consent and ethical committee of Nagasaki University approval was obtained in each case.

Photofrin® (Axcan Pharma, Birmingham, AL, USA) at a dose of 2 mg/kg \[3,5\] was given by intravenous infusion 48 hours prior to surface laser irradiation. An excimer dye laser was used to deliver monochromatic red light at 630 nm. An optical fiber was applied to irradiate the targeted oral lesions. A light dose of 100 - 200 J/cm² was delivered to each area. The patients were counselled to avoid direct sunlight for six weeks following sensitization. Treated areas were biopsied to confirm the effects of PDT 2 to 9 weeks after laser irradiation.

A complete response was defined as the disappearance of all known disease at least 4 weeks after PDT.

Computer-assisted image analysis of cell nuclei

The cell nuclei of lesions were examined with computer-assisted image analysis (Macintosh Image 1.62 on a Power Book G4, Apple Computer, Cupertino, CA, USA) for quantitative measurement of the nuclear areas. Three fields of randomly sampled haematoxylin and eosin stained biopsy specimens were photographed using a Coolpix 4500 digital camera (Nikon, Tokyo, Japan) at 65x magnification. After saving the captured image in a personal computer, 100 nuclei per slide were measured and the nuclear area was calculated (Figure 1). The mean nuclear area (NA) and standard deviation (SD) of NA were calculated. The NA and SD of each specimen were aggregated. The coefficient of variation of the nuclear area (NACV) was calculated as the objective parameter of anisonucleosis by the following formula: NACV (%) = (SD of NA)/(mean NA) x 100 \[13\].

Figure 1. Computer-assisted image analysis of lesion cell nuclei (haematoxylin and eosin stain, original magnification x65).
PCNA immunohistochemistry

Four µm sections were prepared for each specimen, mounted on poly L-lysine-coated glass slides, and dried overnight on a hot plate at 37 °C to promote adhesion. Endogenous peroxidase was inhibited by treatment with 0.3% H₂O₂ in methanol for 30 min. After three washes in phosphate-buffered saline (PBS), the sections were incubated with diluted normal blocking serum for 30 min. Anti-PCNA monoclonal mouse antibody (PC10, 1 : 100 diluted, DAKO M0897, Denmark) was applied for 30 min at room temperature, followed by diluted biotinylated secondary antibody for 30 min, and ABC reagent for 30 min using a Vectastain Elite ABC Kit (Vector Laboratories, Burlingame, CA, USA). PBS was used instead of anti-PCNA antibody as a negative control. The PCNA labelling index (LI) of the lesion cells was determined. Both the labelled and unlabelled lesion cells were counted with the aid of a squared eyepiece graticule (Nikon, Tokyo, Japan) in square fields (0.0625 mm²) at a magnification of 400x. The PCNA LI of the lesion cells was defined as the percentage of PCNA-positive cells in 1000 lesion cells counted from four randomly selected fields.

Statistical analysis

Statistical analysis was performed using the Mann-Whitney U-test for the NA of biopsy specimens between before and after PDT in each case. Paired t-test was also used for statistical comparison of the mean NA and NACV in biopsy specimens before and after PDT in both the nonrecurrent and recurrent group. Differences between values before and after PDT were considered statistically significant if P values were less than or equal to 0.05.

RESULTS

A complete response was achieved in all cases (Figures 2 and 3). Stomatitis was observed for one or two weeks after PDT in all cases. However, other severe adverse side effects were not observed in all cases. Furthermore, no evidence of residual tumour was found in biopsy specimen 4 to 9 weeks after PDT in cases 1 to 13. Case 1, 2, 3-i, 4, 5 and case 17 developed recurrent lesions at the treatment site 4 to 30 months after PDT. These were treated with surgical excision or repeat PDT. The histological diagnosis of each case is shown in Table 1. The mean NAs before PDT were from 54.1 to 158.8 µm² (median = 91.4 µm², SD = 23.0 µm²), and those after PDT were from 24.6 to 106.8 µm² (median = 66.5 µm², SD = 21.7 µm²). Furthermore, medians of mean NAs before PDT were 92.4 µm² (SD = 23.5 µm²) and 88.5 µm² (SD = 23.2 µm²) in nonrecurrent group and recurrent group, respectively. In cases 1 and 3-i, the mean NAs after PDT were significantly larger than those before PDT. In cases 2, 3-ii, 3-iii, 7 and 19, there was no significant difference in mean NA before and after PDT. In cases 4, 5, 6, 8-18 and 20, the mean NAs after PDT were significantly smaller than those before PDT (Table 1). In the nonrecurrent group, the mean NA after PDT was significantly lower than that before PDT (Figure 4A; P = 0.026), although there was no significant difference in mean NA before and after PDT in the recurrent group (Figure 4B; P = 0.5938). The mean NACV before PDT was 36.4% (from 15.8 to 58.1%, SD = 9.6%), and that after PDT was 31.3% (from 19.5 to 54.3%, SD = 9.7%) in the nonrecurrent group (Figure 5A), while that before PDT was 32.0% (from 19.9 to 38.6%, SD = 7.0%) and that after PDT was 28.6% (from 22.8 to 34.8%, SD = 4.2%) in the recurrent group (Figure 5B).
Table 1. Nuclear changes of oral lesions epithelial cells before and after photodynamic therapy

<table>
<thead>
<tr>
<th>Case (Age, Sex)</th>
<th>Site of lesions (TNM classification)</th>
<th>Histological diagnosis (pre-PDT/post-PDT)</th>
<th>Mean of NA (μm²; pre-PDT/post-PDT)</th>
<th>NACV (%) pre-PDT/post-PDT</th>
<th>PCNA LI (%) pre-PDT/post-PDT</th>
<th>Follow-up or recurrent term</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 (56, F)</td>
<td>tongue (T2N0M0)</td>
<td>SCC/ED, moderate</td>
<td>57.7/82.2</td>
<td>28.4/27.0</td>
<td>20.5/20.0</td>
<td>30 months +</td>
</tr>
<tr>
<td>2 (90, F)</td>
<td>buccal mucosa (T2N0M0)</td>
<td>SCC/ED, moderate</td>
<td>105.2/106.8</td>
<td>19.9/27.4</td>
<td>17.2/16.7</td>
<td>15 months +</td>
</tr>
<tr>
<td>3 (69, F)</td>
<td>i) lt. buccal mucosa (T1N0M0)</td>
<td>SCC/ED, severe</td>
<td>60.4/97.0</td>
<td>32.5/22.8</td>
<td>36.3/20.7</td>
<td>11 months +</td>
</tr>
<tr>
<td>4 (83, M)</td>
<td>ii) rt. lower gingiva (T1N0M0)</td>
<td>SCC/gingivitis</td>
<td>93.7/96.5</td>
<td>36.2/54.3</td>
<td>27.3/25.1</td>
<td>37 months -</td>
</tr>
<tr>
<td></td>
<td>iii) rt. buccal mucosa (T1N0M0)</td>
<td>SCC/ED</td>
<td>90.2/85.4</td>
<td>44.6/26.6</td>
<td>22.7/7.7</td>
<td>37 months -</td>
</tr>
<tr>
<td>5 (70, M)</td>
<td>hard palate (T2N0M0)</td>
<td>SCC/stomatitis</td>
<td>108.8/84.3</td>
<td>38.6/27.4</td>
<td>44.0/21.1</td>
<td>15 months +</td>
</tr>
<tr>
<td>6 (63, F)</td>
<td>oral floor (T2N0M0)</td>
<td>SCC/stomatitis</td>
<td>95.2/65.8</td>
<td>37.3/32.1</td>
<td>22.5/10.8</td>
<td>4 months +</td>
</tr>
<tr>
<td>7 (80, F)</td>
<td>hard palate (T1N0M0)</td>
<td>SCC/gingivitis with moderate ED</td>
<td>127.2/24.6</td>
<td>43.9/43.6</td>
<td>16.0/21.0</td>
<td>20 months -</td>
</tr>
<tr>
<td>8 (85, F)</td>
<td>upper gingival (T2N0M0)</td>
<td>SCC/gingivitis</td>
<td>95.6/63.1</td>
<td>34.2/25.8</td>
<td>52.7/17.0</td>
<td>22 months -</td>
</tr>
<tr>
<td>9 (75, M)</td>
<td>buccal mucosa (T1N0M0)</td>
<td>SCC/sialadenitis</td>
<td>127.2/24.6</td>
<td>43.9/43.6</td>
<td>16.0/21.0</td>
<td>20 months -</td>
</tr>
<tr>
<td>10 (74, F)</td>
<td>lower gingival (T1N0M0)</td>
<td>SCC/gingivitis</td>
<td>82.3/53.8</td>
<td>32.1/29.9</td>
<td>12.5/3.8</td>
<td>15 months -</td>
</tr>
<tr>
<td>11 (78, F)</td>
<td>lower gingival (T1N0M0)</td>
<td>SCC/gingivitis</td>
<td>97.8/59.7</td>
<td>58.1/40.4</td>
<td>19.6/11.9</td>
<td>18 months -</td>
</tr>
<tr>
<td>12 (76, M)</td>
<td>Tongue (T2N0M0)</td>
<td>SCC/glossitis</td>
<td>96.0/81.1</td>
<td>39.7/27.5</td>
<td>25.0/5.3</td>
<td>18 months -</td>
</tr>
<tr>
<td>13 (66, M)</td>
<td>lower gingival (T3N0M0)</td>
<td>verrucous carcinoma/ED, severe</td>
<td>158.8/35.0</td>
<td>41.6/35.3</td>
<td>26.7/16.4</td>
<td>10 months -</td>
</tr>
<tr>
<td>14 (72, M)</td>
<td>tongue</td>
<td>ED, moderate/ED, mild</td>
<td>54.1/41.6</td>
<td>35.9/32.0</td>
<td>49.9/41.7</td>
<td>19 months -</td>
</tr>
<tr>
<td>15 (29, F)</td>
<td>tongue</td>
<td>ED, mild/glossitis</td>
<td>87.1/71.2</td>
<td>27.2/39.3</td>
<td>17.1/22.0</td>
<td>19 months -</td>
</tr>
<tr>
<td>16 (79, F)</td>
<td>hard palate</td>
<td>ED, mild /ED, mild</td>
<td>81.9/51.7</td>
<td>15.8/21.0</td>
<td>25.3/17.9</td>
<td>30 months -</td>
</tr>
<tr>
<td>17 (65, F)</td>
<td>tongue</td>
<td>ED, severe/glossitis</td>
<td>103.5/43.7</td>
<td>35.6/34.8</td>
<td>19.1/13.7</td>
<td>30 months +</td>
</tr>
<tr>
<td>18 (61, M)</td>
<td>soft palate</td>
<td>ED/stomatitis with mild ED</td>
<td>82.6/59.7</td>
<td>39.6/23.9</td>
<td>43.5/37.5</td>
<td>33 months -</td>
</tr>
<tr>
<td>19 (70, F)</td>
<td>buccal mucosa</td>
<td>ED, moderate/ED, mild</td>
<td>82.2/80.2</td>
<td>26.2/22.2</td>
<td>35.4/37.1</td>
<td>16 months -</td>
</tr>
<tr>
<td>20 (67, M)</td>
<td>tongue</td>
<td>ED, mild/glossitis</td>
<td>95.6/74.0</td>
<td>31.7/22.5</td>
<td>35.0/47.0</td>
<td>7 months -</td>
</tr>
</tbody>
</table>

*Existence of statistically significant differences, P < 0.0001.

†Existence of statistically significant differences, P < 0.0243.

F = female; M = male; SCC = squamous cell carcinoma; ED = epithelial dysplasia; NA = nuclear area; NACV = coefficient of variation of the nuclear area; PCNA LI = proliferating cell nuclear antigen labelling index; PDT = photodynamic therapy.

Figure 4A. Mean nuclear area (NA) in nonrecurrent group. There was significant difference in mean NA before and after PDT (P = 0.0026).

Figure 4B. Mean nuclear area (NA) in recurrent group. There was no significant difference in mean NA before and after PDT (P = 0.5983).
Figure 5A. Mean coefficient of variation of the nuclear area (NACV) in nonrecurrent group. There was no significant difference in mean NACV before and after PDT (P = 0.0920).

Figure 5B. Mean coefficient of variation of the nuclear area (NACV) in recurrent group. There was no significant difference in mean NACV before and after PDT (P = 0.2689).

The mean NACV after PDT tended to be lower than that before PDT in the nonrecurrent group, although the difference was not significant (P = 0.0920).

PCNA-positive cells were distinguished as the brown-stained nuclei in immunohistochemical staining. The mean PCNA LI before PDT was 30.3% (from 12.5 to 52.7%, SD = 12.3%), and that after PDT was 22.5% (from 3.8 to 47.0%, SD = 13.0%) in the nonrecurrent group (Figure 6A), while that before PDT was 24.3% (from 17.2 to 44.0%, SD = 10.9%) and that after PDT was 15.0% (from 10.8 to 21.1%, SD = 4.2%) in the recurrent group (Figure 6B). There were significant differences in PCNA LI values before and after PDT in both the nonrecurrent and recurrent group (P = 0.0188 in nonrecurrent group, P = 0.0496 in recurrent group).

DISCUSSION

Morphological change in the nuclei is a useful marker for assessing the effects of radiotherapy [14] and chemotherapy [15,16]. With regard to specific PDT effects, plasma membranes are the most important targets [17,18], and mitochondria are also critical targets [19,20]. In our previous study, we also clarified...
that the morphological change of mouse tumour cell nuclei subjected to PDT was available to assess the anti-tumour effects of PDT [11]. In the present human study, the mean NA after PDT significantly decreased compared to that before PDT in the non-recurrent group while this difference was not significant in the recurrent group. PDT induces DNA fragmentation in some cell lines [21], and may induce the reduction of cell nuclei after PDT. Accordingly, it was speculated that when PDT targets cells whose nuclei are barely susceptible to PDT, there is a risk of recurrence. NACV is also useful for objectively evaluating nuclear pleomorphism [13-22]. The significant reductions of NACV reflect the morphologic alterations of tumour nuclei with narrow dispersion of their size, and the value increases with the malignant potential of the lesion [13]. In a previous study, we experimentally demonstrated that the reduction of NACV reflects the anti-tumour effects of PDT to some extent [11]. However, in this study there was no significant difference in mean NACV before and after PDT in either group, although NACV after PDT was lower than that before PDT in the non-recurrent group (P = 0.0920). On the other hand, the mean PCNA LI after PDT was significantly lower than that before PDT in the both the non-recurrent and recurrent group. PCNA synthesis occurs in the latter part of G1 and throughout the S-phase, predominantly increasing in the S-phase [23], while the photosensitivity of cells increases during the G1 to mid-S-phase [1]. Accordingly, the significant decrease of PCNA LI after PDT may have resulted from susceptibility to PDT in the PCNA-positive tumour cells. These results are likely to be indicative that NACV and PCNA LI are not predictive markers for tumour recurrence after PDT. The primary advantage of PDT is that minimal damage is caused to healthy tissue, and PDT can be applied repeatedly at the same site because the laser used in PDT is nonthermal. However, targets are generally superficial or early-stage lesions [24]. In order to apply PDT to advanced lesions, or to have a better prognosis, fractionated laser irradiation [25,26] and adjuvant chemotherapies [26,27] should be made available. Furthermore, secondary PDT is likely to be indicated in cases which have no significantly reduced mean NA in the biopsy specimen after PDT, even if there is no evidence of residual tumour.

CONCLUSIONS

The non-reduction of nuclear area after photodynamic therapy may predict a possibility of recurrence of oral malignancy and epithelial dysplasia, although further investigations on more examples will be required to precisely predict the recurrence of oral malignancy and epithelial dysplasia.

ACKNOWLEDGMENTS AND DISCLOSURE STATEMENTS

The authors report no conflicts of interest related to this study.

REFERENCES


